

## Studies of positronium atoms in normal and cancer tissues and cultured cell lines - biomedical application

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New Quantum Horizons: From Foundations To Biology Symposium Frascati





EST 1364



## 1) Motivation

- 2) Cancer vs. normal cells
- 3) PALS studies of tumor and normal tissues in vitro
- 4) First PALS studies of human tissues in vitro with J-PET

Outline

- 5) PALS studies of cells cultures in vitro
- 6) Summary and future plans



## Motivation

 $\rightarrow$  Determination of early and advanced stages of carcinogenesis by observing changes in biomechanical parameters between normal and cancer cells

 $\rightarrow$  PALS parameters (lifetime, intensity, radius) are related with temporal dynamics of nanostructures in whole cells and tissues

 $\rightarrow$  Combining J-PET scanner with PALS technique – better diagnostic tool

 $\rightarrow$  Positronium imaging in the human body in vivo !

 $\rightarrow$  Over 50 publication and 15 patents on J-PET and positronium imaging

P. Moskal et al., Feasibility study of the positronium imaging with the J-PET tomograph, submitted to Physics in Med. And Bio.

P. Moskal, TOF-PET tomograph and a method of imaging using a TOF-PET tomograph, based on a probability of production and lifetime of a positronium, patent no. P405185, PCT/EP2014/068374



## Cancer vs normal cells Normal Cancer Cytoplasm Nucleus -Nucleolus -Chromatin- $\rightarrow$ Large cytoplasm $\rightarrow$ Small cytoplasm $\rightarrow$ Single nucleus and nucleolus $\rightarrow$ Multiple and large nucleus and nucleolus $\rightarrow$ Fine chromatin $\rightarrow$ Coarse chromatin $\rightarrow$ Smaller number of dividing cells $\rightarrow$ Large number of dividing cells $\rightarrow$ Similar in shape and size $\rightarrow$ Variation in cells shape and size

- $\rightarrow$  Disorganized arrangement of cells
- $\rightarrow$  Immortal

### en.uj.edu.pl

https://visualsonline.cancer.gov/details.cfm?imageid=2512

 $\rightarrow$  Organized arrangement of cells

 $\rightarrow$  Apoptosis



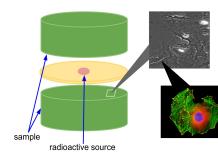
## PALS setup

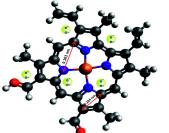
 $\rightarrow$  Two BaF<sub>2</sub> detectors with resolution ~250 ps

 $\rightarrow$   $^{22}Na$  source in Kapton foil with activity  $\sim$  1 MBq sandwich between sample

 $\rightarrow$  PALS spectra analysis with PALS\_Avalanche program developed by K. Dulski – J-PET collaboration

K. Dulski et. al., Analysis procedure of the positronium lifetime spectra for the J-PET detector, Acta Phys. Polon. B48 no. 10, 1611 (2017)

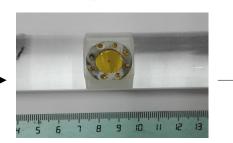


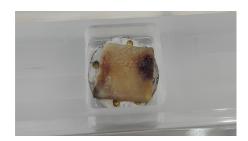


e⁺ + e
C
Fe²+
N
H

Heme group in hemoglobin molecule

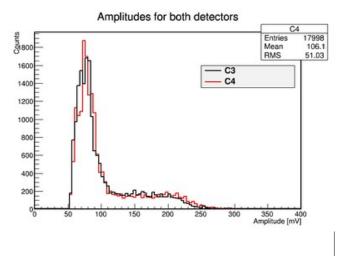


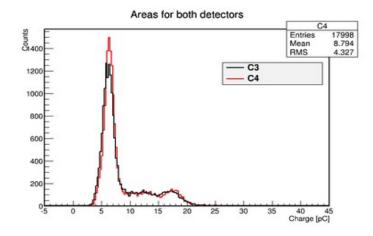




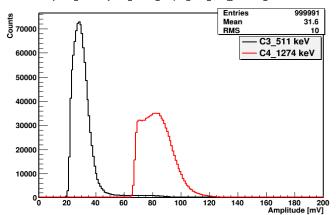


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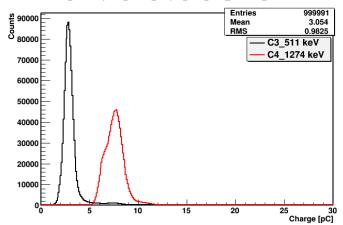




Amplitude\_CardiacMyxoma\_Tumor1\_Sample1\_Fixed\_1MBq\_BioHolder\_10112018.dat



Area\_CardiacMyxoma\_Tumor1\_Sample1\_Fixed\_1MBq\_BioHolder\_10112018.dat

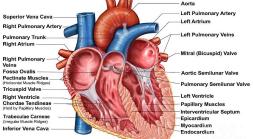




Cardiac Myxoma

 $\rightarrow$  primitive connective tissue tumor (benign), very rare in comparison to metastatic tumors

- $\rightarrow$  75 % of them are located in the left atrium
- $\rightarrow$  occur mainly in people over the age of 50



Types	Solid	Papillary
Surface	smooth	irregular
Mass	firm	soft, gelatinous
Calcification	+	-
Embolism	-	+

### Fixed in formaldehyde:

1) Myxoma I (6 samples for study, around 2 mm thick) 72 years old women

- 2) Myxoma II (1 sample) 61 year old men
- 3) Myxoma III (3 samples) 59 year old men
- 4) Myxoma IV (3 samples) 54 year old woman

### Not fixed (fresh):

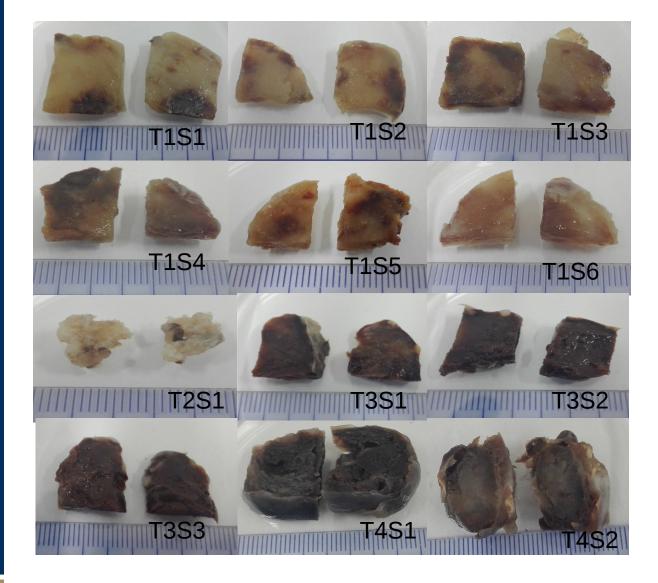
5) Myxoma V (1 sample) 77 year old men – measured within 4 hours after the surgery

https://healthjade.com/human-heart-health/

## Cardiac Myxoma



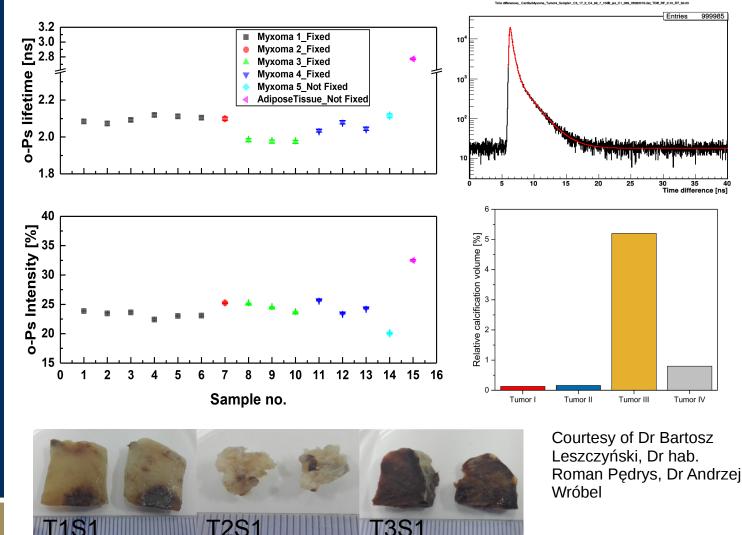
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## Cardiac Myxoma - PALS

- $\rightarrow$  Samples fixed in formaldehyde  $\rightarrow$  not decomposing/changing in time
- $\rightarrow$  Fresh sample measured within 4 hours after surgery
- $\rightarrow$  Time of measurement ~60 min  $\rightarrow$  1 mln counts

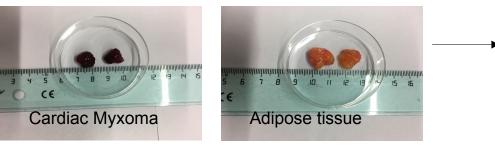




# Cardiac Myxoma – JPET vs PALS

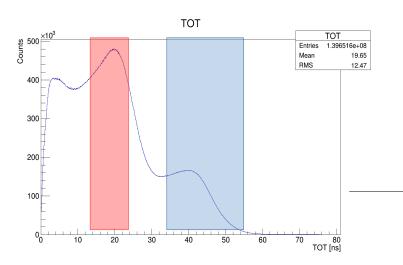
 $\rightarrow$  Samples after extraction from patient were placed in sterile container with DMEM medium supplemented with 10% FBS, Penicillin/Streptomycin and HEPES buffer

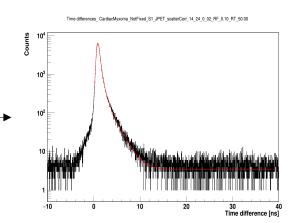
- $\rightarrow$  Fresh sample measured within 4 hours after surgery
- $\rightarrow$  Time of measurement ~70-80 min  $\rightarrow$  1 mln counts





### $\rightarrow$ Cut on TOT 14-24 ns (511 keV) and 35-55 ns (1274 keV)



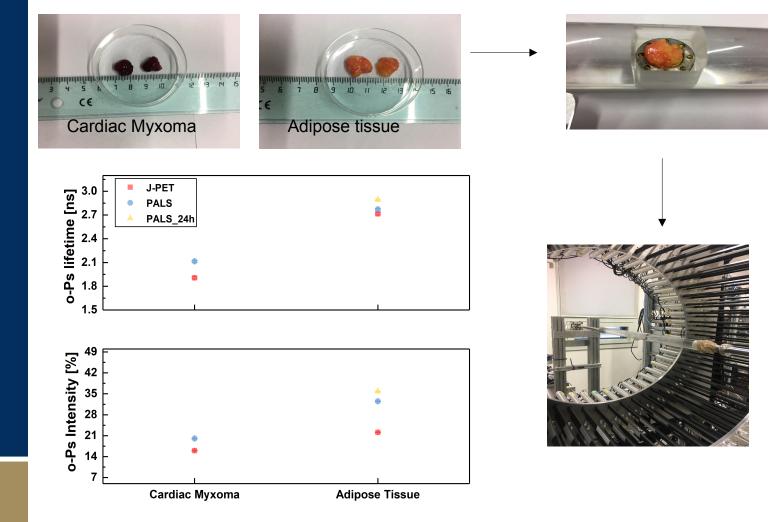




# Cardiac Myxoma – JPET vs PALS

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# PALS – Cells culture in vitro

### Human cell lines:

1) Melanocytes HEMa-LP from ThermoFisher

- 2) Melanoma WM115 from ATCC
- 3) Melanoma WM266 from ATCC

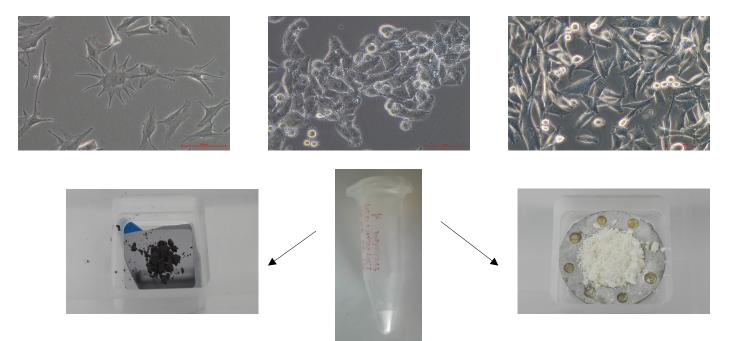
→ Cells were cultured in M254/RPMI 1640 medium supplemented with 4.5g/L glucose, 2 mM L-glutamine and HGMS-2/10% fetal bovine serum, additionally Penicillin 100U/ml and Streptomycin 100 ug/ml was added to the culture.

 $\rightarrow$  Medium was changed every 2 days.

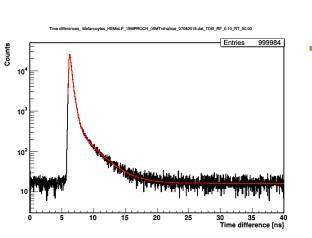
 $\rightarrow$  Culture was incubated at 37°C in 5% CO<sub>2</sub> humidified atmosphere rinse with PBS

w/o Ca2+, Mg2+ and passage with 0.25% Trypsin every 3-4 days.

 $\rightarrow$  Each samples contains cells from 8 T75 flasks, harvest upon 100% confluation and freeze – dried (lyophilized) – In total ~ 20\*10<sup>7</sup> cells (~0.5 cm<sup>3</sup>)



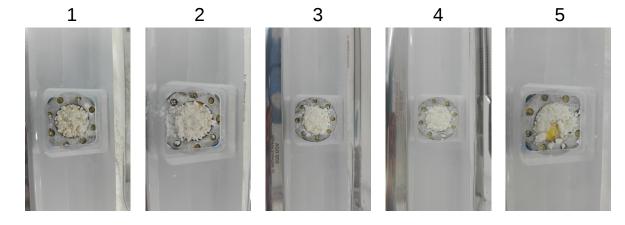




## PALS – Cells culture in vitro

### Human cell lines:

- 1) Melanocytes HEMa-LP from ThermoFisher
- 2) Melanoma WM115 from ATCC
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### Freeze Mediums:

0

- 1) M254/RPMI 1640+ P/S+ 20% FBS + 10% DMSO
- 2) 10% DMSO + PBS w/o <sup>2+</sup>Ca, <sup>2+</sup>Mg
- 3) PBS w/o <sup>2+</sup>Ca, <sup>2+</sup>Mg
- 4) 1.5 M PROH( propylene glycol) + 0.5 M D-trehalose in PBS w/o  $^{2+}$ Ca,  $^{2+}$ Mg
- 5) 0.25 M D-trehalose in PBS w/o <sup>2+</sup>Ca, <sup>2+</sup>Mg



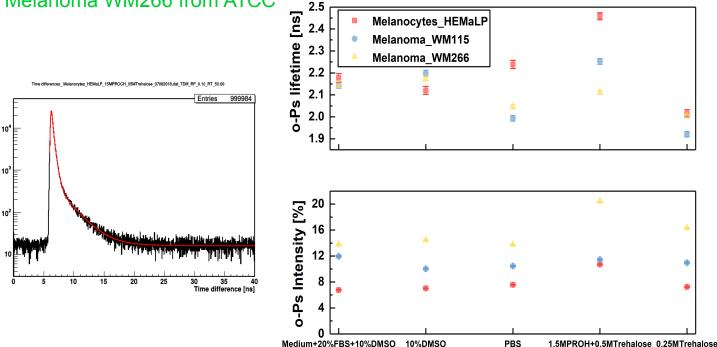
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### Human cell lines:

Counts

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## PALS – Cells culture in vitro

s-P

Medium+20%FBS+10%DMSO

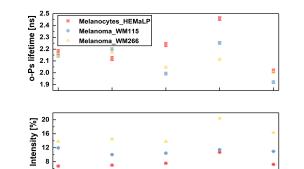


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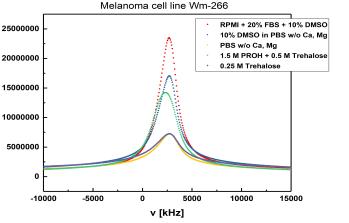
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PBS

1.5MPROH+0.5MTrehalose 0.25MTrehalose

	Viability [%]	% of water	Remaining mass [%]
1	43.8	44	-
2	40.5	31	14.69
3	5.4	22	14.24
4	49.7	49	46.25
5	10.7	16	24.62



10%DMSO

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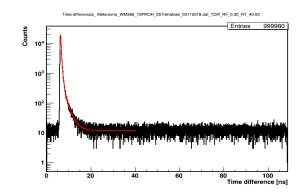






	Viability before [%]	Viability after [%]
1	97.4	95.8
2	96.8	96.7
3	96.6	97.3
4	97.9	97.7
5	98.1	97.6
6	98.2	98.0
7	97.1	91.4

Sample	o-Ps lifetime[ns]	o-Ps Intensity [%]
1	2.01(01)	15.77(09)
2	2.01(01)	18.16(09)
3	2.06(01)	17.23(09)
4	1.99(01)	17.87(09)
5	2.05(01)	16.94(09)
6	1.96(01)	17.17(10)
7	2.00(01)	18.23(10)
7_2h	2.01(01)	18.17(09)





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# Summary and future plan

 $\rightarrow$  PALS is applicable to study biological structures

 $\rightarrow$  Preliminary results shown that PALS parameters differ for normal and cancer cells and tissue

 $\rightarrow$  First studies of human tissue on JPET scanner proves that o-Ps lifetime can be used as additional diagnostic parameter

 $\rightarrow$  Development of the method for sample preparation in order to study alive cell cultures

- $\rightarrow$  Studies with alive cell cultures and tissues comparing normal vs cancer
- $\rightarrow$  Primary cell culture derived from cardiac myxoma tumor
- $\rightarrow$  Checking for possible o-Ps formation model in living cells

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### Thank you for your attention

